

## Correction

### ENGINEERING

Correction for “Computer simulations suggest that prostate enlargement due to benign prostatic hyperplasia mechanically impedes prostate cancer growth,” by Guillermo Lorenzo, Thomas J. R. Hughes, Pablo Dominguez-Frojan, Alessandro Reali, and Hector Gomez, which was first published January 7, 2019; 10.1073/pnas.1815735116 (*Proc. Natl. Acad. Sci. U.S.A.* **116**, 1152–1161).

The authors note that Eq. 1 on page 1158 appeared incorrectly. The corrected equation appears below.

$$\frac{\partial \phi}{\partial t} = \nabla \cdot (MD_{\phi} \nabla \phi) + M \left( \chi s - A\phi - \frac{1}{\tau} \frac{dF(\phi)}{d\phi} \right)$$

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Published online June 10, 2019.

[www.pnas.org/cgi/doi/10.1073/pnas.1908514116](http://www.pnas.org/cgi/doi/10.1073/pnas.1908514116)



# Computer simulations suggest that prostate enlargement due to benign prostatic hyperplasia mechanically impedes prostate cancer growth

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**Prostate cancer and benign prostatic hyperplasia are common genitourinary diseases in aging men. Both pathologies may coexist and share numerous similarities, which have suggested several connections or some interplay between them. However, solid evidence confirming their existence is lacking. Recent studies on extensive series of prostatectomy specimens have shown that tumors originating in larger prostates present favorable pathological features. Hence, large prostates may exert a protective effect against prostate cancer. In this work, we propose a mechanical explanation for this phenomenon. The mechanical stress fields that originate as tumors enlarge have been shown to slow down their dynamics. Benign prostatic hyperplasia contributes to these mechanical stress fields, hence further restraining prostate cancer growth. We derived a tissue-scale, patient-specific mechanically coupled mathematical model to qualitatively investigate the mechanical interaction of prostate cancer and benign prostatic hyperplasia. This model was calibrated by studying the deformation caused by each disease independently. Our simulations show that a history of benign prostatic hyperplasia creates mechanical stress fields in the prostate that impede prostatic tumor growth and limit its invasiveness. The technology presented herein may assist physicians in the clinical management of benign prostate hyperplasia and prostate cancer by predicting pathological outcomes on a tissue-scale, patient-specific basis.**

prostate cancer | benign prostatic hyperplasia | mathematical oncology | patient-specific | isogeometric analysis

Despite its small size and supporting role in the male genitourinary system, the prostate is the site of two major pathologies in older men: prostate cancer (PCa) and benign prostatic hyperplasia (BPH) (1). PCa is almost always an adenocarcinoma, a form of cancer that originates in epithelial tissues with glandular organization, for instance, the prostatic tissue in charge of producing certain substances of semen. Currently, PCa is the second most common cancer and the fifth leading cause of death from cancer in men worldwide, with an estimated 1.1 million new cases and 307,000 deaths in 2012 (2). PCa is usually diagnosed and treated when it is still localized within the prostate (1). The majority of PCa cases originate in the peripheral zone (PZ) of the prostate (70%), but PCa can also arise in the transition zone (20%) and in the central zone (10%). BPH consists of the pathological enlargement of the prostate with age, with a prevalence increasing from 50% in men in their 50s to about 70% in men in their 70s (3). This condition may arise in the transition zone or the periurethral glands (1). The growing tissue may obstruct urinary and ejaculatory flow, hence causing bothersome lower-urinary tract symptoms. As BPH does not develop in the PZ, the glandular prostate is segmented into two zones in radiological studies: the PZ and the central gland (CG), which contains the central, transition, and periurethral zones (1, 4, 5). The CG has a larger and denser stromal component that

becomes more compact during BPH, which results in an overall lower signal intensity in T2-weighted magnetic resonance (MR) images. However, the signal intensity of the CG is usually rather heterogeneous due to the varying proportions of stromal and glandular hyperplasia (1, 4–7).

Beyond anatomical location, BPH and PCa possess other similarities (1, 8, 9). At the epidemiological level, both diseases affect older men, have increasing incidence with age, and frequently coexist in the same patient. These pathologies may induce lower-urinary tract symptoms and increase serum levels of prostatic specific antigen (PSA), a prostate activity biomarker used in PCa diagnosis and staging. Additionally, PCa and BPH share some genetic alterations, are hormone dependent, have been correlated with prior inflammation, and might be part of the metabolic syndrome. Despite the evidence suggesting links between PCa and BPH, the relationship between these diseases has been under continuous debate in the medical community due to the existence of contradictory studies and the scarcity of research thoroughly confirming the proposed hypothetical links (8, 9). Gaining knowledge about the connection between PCa and BPH would challenge the current clinical standards and potentially enhance treatment and prevention of both diseases.

## Significance

**Benign prostatic hyperplasia (BPH) is a common disease in aging men that causes the prostate to enlarge progressively. Men with larger prostates tend to harbor prostatic tumors with more favorable features. The underlying mechanisms that explain this interaction between BPH and prostate cancer (PCa) are largely unknown. Here, we propose that BPH may mechanically impede PCa growth by producing increasingly intense mechanical stresses in the prostate over time, which are known to slow down tumor dynamics. To explore this hypothesis, we ran a qualitative simulation study using an extension of our mathematical model of PCa growth including the mechanical deformation of the prostate under BPH and PCa. The proposed mechanism suggests relevant shifts in clinical management of PCa and BPH.**

Author contributions: G.L., T.J.R.H., A.R., and H.G. designed research; G.L., T.J.R.H., A.R., and H.G. performed research; G.L., P.D.-F., and H.G. contributed new reagents/analytic tools; G.L., T.J.R.H., A.R., and H.G. analyzed data; and G.L., T.J.R.H., A.R., and H.G. wrote the paper.

Conflict of interest statement: H.G., T.J.R.H., and G.L. are listed as coinventors in a patent application that has been filed by The University of Texas at Austin. The invention leverages part of the technology presented in this paper.

Reviewers: K.G., University of Michigan; and E.K., Stanford University.

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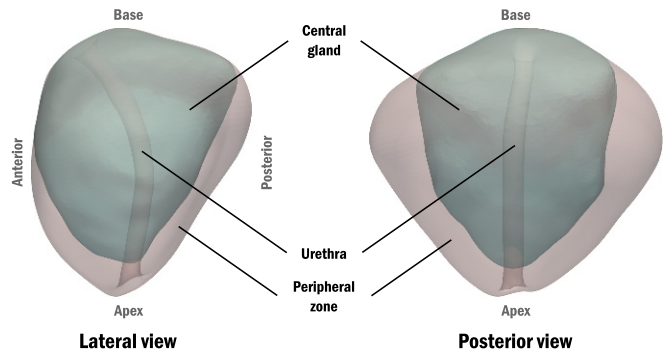
This article contains supporting information online at [www.pnas.org/lookup/suppl/doi:10.1073/pnas.1815735116/-DCSupplemental](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1815735116/-DCSupplemental).

Published online January 7, 2019.

The biopsy detection rate of PCa in large prostates is known to be lower than in small prostates (10–12). This low detection rate has been attributed to various artifacts, such as sampling errors and a biopsy selection bias in patients with increasing PSA, possibly caused by BPH (10, 13, 14). However, it appears that neither increasing sampling nor using more advanced medical imaging to perform biopsy has leveled the detection rate across prostate volume groups (11, 12, 15, 16). Recent studies on large series of radical prostatectomy specimens show that tumors arising in larger prostates present favorable pathological features, such as lower aggressiveness measured with the Gleason score (1), smaller tumor volume, and lower risk of extraprostatic extension or seminal vesicle invasion (10, 17–21). These studies provide solid evidence to suggest that large prostates may exert a protective effect against PCa, but the underlying mechanisms are not known.

Here, we propose a mechanical explanation for this phenomenon: BPH exerts an inhibitory effect on PCa growth due to the accumulation of mechanical stress. The rapid overproliferation of cancerous cells leads to high cell densities in their confined microenvironment within the harboring tissue. As a result, tumors develop a high compressive hydrostatic stress state in their interiors and exert outward forces as they grow. These forces deform the neighboring tissues, generating stress fields aimed at resisting the mechanical load imposed by tumor growth. This phenomenon is known as the tumor mass effect (22–28). In turn, these mechanical stress fields are known to exert an inhibitory effect on tumor growth both directly, by slowing down tumor dynamics, and indirectly, by deforming and collapsing the local vasculature in the tumor region. BPH creates a compressive hydrostatic stress state within the CG and outward forces that deform the PZ (1, 4, 5). Additionally, the confinement of the prostate in the pelvic area further contributes to the internal mechanical stress fields because the surrounding tissues resist the pathological deformation of this organ. Here, we show that, as the CG enlarges over time, the mechanical stress fields created by BPH intensify and reach larger values than those created by the tumor itself. Hence, BPH may also exert an inhibitory effect on PCa growth but its impact has been overlooked. Indeed, if clinically validated, this mechanism may produce a major change in BPH treatment. The current paradigm aims at reducing the CG volume to alleviate BPH symptoms via surgery or medication (1). According to the proposed mechanism, this would reduce the mechanical stresses in the prostate and hence promote the growth of coexisting PCa. This would put the patient in a dangerous predictive scenario, especially if the tumor was not previously diagnosed.

To analyze whether enlarged prostates may provide protection against PCa growth, we need to determine whether the stress induced by BPH is enough to significantly produce an inhibitory effect on PCa. This fundamentally requires a 3D, anatomically faithful, mechanical model of the prostate. The amount by which the growth rate of the tumor decreases with stress may be estimated from experiments in the literature. To perform this study, we derive a tissue-scale, patient-specific mechanically coupled mathematical model for PCa growth and perform a series of computer simulations. The study and design of mathematical models to forecast the evolution of tumors using computer simulations have given rise to the field of mathematical oncology (29–31). These models aim at complementing the current clinical practice in oncology by assisting physicians in better estimating disease progression and designing optimal treatment schemes. In particular, several mathematical models have incorporated the tumor mass effect to improve the prediction of the growth of various types of cancers (32–38). Fig. 1 depicts the geometry of the prostate used in our simulations, which we extracted from the MR images and anatomical segmentations of a patient suffering from PCa and BPH.



**Fig. 1.** Patient-specific local anatomy of the prostate. From a radiological perspective, the prostate is divided into central gland (CG) and peripheral zone (PZ). BPH takes place in the CG and most cases of PCa arise in the PZ. We extracted the geometry of the patient's prostate and CG from their corresponding segmentations provided on axial T2-weighted MR images. The volumes of the prostate, CG, and PZ at MRI date are 52.81 cc, 33.15 cc, and 19.66 cc, respectively. The major diameters of the prostate at MRI date have a length of 53.49 mm, 38.35 mm, and 52.01 mm in lateral, anteroposterior, and craniocaudal directions, respectively.

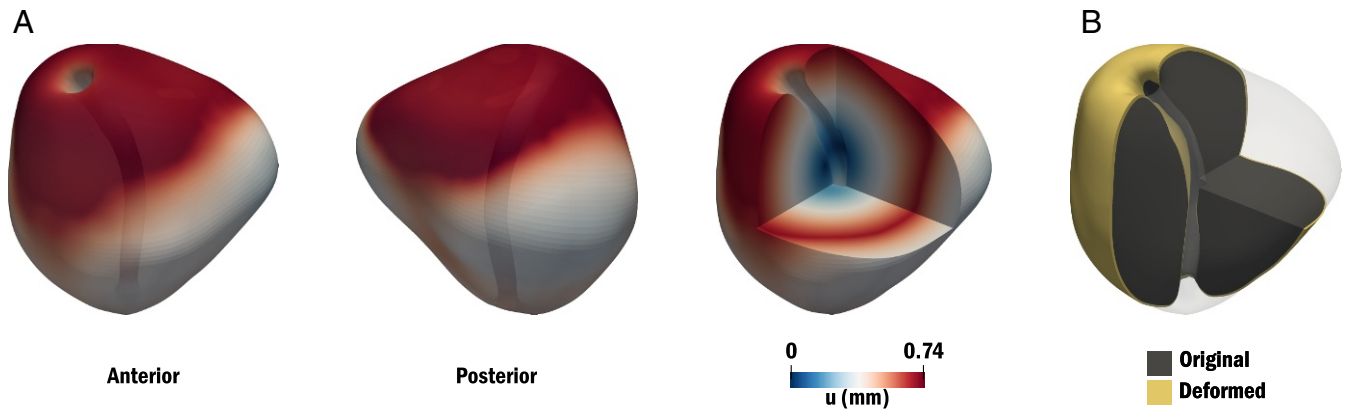
In this work, we extend our previous model of organ-confined PCa growth (39, 40) to include the equations of mechanical equilibrium and define the coupling terms between them and tumor dynamics. As organ-confined PCa growth can be seen as an evolving interface problem, we leverage the phase-field method (41) to account for the coupled dynamics of healthy and tumoral tissue. Our model also estimates PSA dynamics at tissue level by computing the serum concentrations of this biomarker produced at healthy and cancerous regions per unit volume of prostatic tissue (39). Following previous mechanically coupled approaches (32–38), we assume that the deformation of the prostate is a quasistatic phenomenon and we model prostatic tissue as a linear elastic, heterogeneous, isotropic material. As CG is normally stiffer than PZ in older men, we set a higher Young modulus in the CG (1, 4–7, 42–45). The confinement of the prostate in the pelvic region is modeled with Winkler-inspired boundary conditions on the external surface of the organ, while free displacement is imposed along the urethra. BPH and PCa are modeled as pressure terms in the constitutive equation of the prostatic tissue acting within the CG and the tumor, respectively. Finally, the inhibitory effect of the mechanical stress fields is estimated with a global factor that slows down tumor dynamics. This coefficient depends on a measure of the loading conditions and the stored elastic energy, so we choose a combination of the Von Mises stress and the hydrostatic stress for this purpose (46). The range of values of this inhibitory factor is adjusted to match the experimental and clinical observations in previous studies of tumor growth (23–28, 32–35). See *Materials and Methods* for more details on the model.

## Results

**Deformation of the Prostate Due to BPH.** Our model predicted that BPH produces the volumetric expansion of the prostate in the perpendicular direction to the CG border, as depicted in Fig. 2 (*SI Appendix, Fig. S1*). The PZ was pushed outward from the borders of the growing CG against the prostate external surface and experienced a very slight shrinkage, as the boundary conditions partially enabled outward displacement.

The extremal values of the displacements were attained at the CG borders that are closer to the external surface of the prostate. The maximum total displacement was 0.74 mm. The urethra was displaced posteriorly and its diameter was virtually unaltered.

The hydrostatic stress was compressive within the CG (−0.20 kPa to −0.13 kPa) and negligible within the PZ (*SI*



**Fig. 2.** Deformation of the prostate caused by BPH over 1 y. (A) Length of the displacement field vector over original anatomy at  $t = 1$  y. (B) Original and deformed geometries of the prostate at  $t = 1$  y.

*Appendix, Fig. S1*). Positive hydrostatic stress appeared in thin PZ regions between the CG border and the prostate boundary, where tension accumulated as the CG expanded. The Von Mises stress was negligible within the CG, except along the urethra (0.27–0.53 kPa). Within the PZ, the Von Mises stress peaked near the borders of the CG (0.20–0.33 kPa) and decreased toward the external surface of the prostate (0.07–0.17 kPa), depending on the distance between them (*SI Appendix, Fig. S1*).

**Deformation of the Prostate Due to PCa.** We considered three artificial tumors placed in characteristic locations within the prostate: the basal PZ, the apical PZ, and the CG. Each tumor had ellipsoidal geometry with one semiaxis measuring 4 mm and the others 3 mm. The largest dimension was oriented in anteroposterior direction. We selected the parameters that regulate tumor dynamics so that these tumors were aggressive and showed the typical morphologies of localized PCa: massive and fingered (39). Fig. 3 shows the growth of these artificial tumors and how they deformed the patient’s prostate (*SI Appendix, Fig. S2*).

All tumors started growing with the massive morphology, which is characteristic of small prostatic cancers. The PZ tumors soon adjusted their geometry to the anatomy of the patient’s prostate boundary (Fig. 3 *A1* and *A2* at  $t = 0.3$  y). Due to the reduced thickness of the PZ around the CG in the patient’s prostate, these PZ tumors invaded the CG early ( $t < 0.1$  y). The CG tumor took longer to reach the PZ and invade it ( $t \approx 0.3$  y). As every tumor grew in size, the intratumoral nutrient concentration decreased. This shortage cued a shape instability that progressively adjusted the tumors to a fingered or lobular morphology, hence ensuring a spatial distribution of nutrient that sustained tumor growth (39). This phenomenon happened by  $t = 0.4$  y for the tumor in basal PZ, by  $t = 0.35$  y for the tumor in apical PZ, and by  $t = 0.55$  y for the CG tumor. The shift in morphology arrested tumor growth momentarily ( $< 0.15$  y) and even reduced the CG tumor volume by 12.2%. However, tumors grew faster and more extensively after this phenomenon.

The CG tumor grew faster and larger than the PZ tumors, whose growth rates and initial volumes were similar. Because the tumor in apical PZ underwent the change in morphology earlier, its volume was larger than that of the basal PZ tumor for the second half of the simulation. The final volumes of the basal PZ tumor, the apical PZ tumor, and the CG tumor were 5.43 cc, 6.82 cc, and 9.02 cc, respectively.

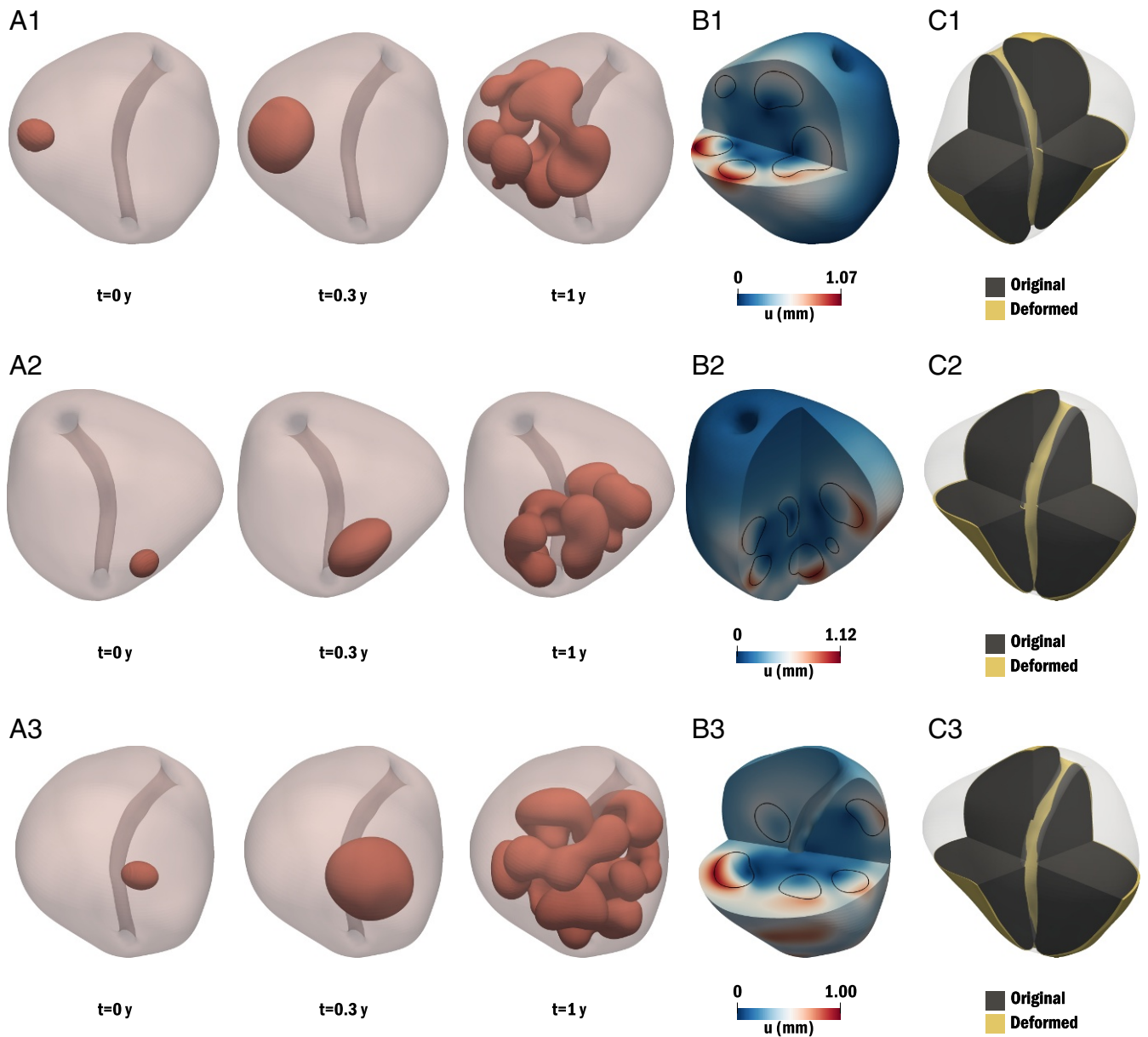
Tumors produced a local swelling deformation. The outermost tumoral structures produced the greatest displacements, which ranged between 0.65 mm and 1.15 mm and created noticeable

smooth lumps on the external prostatic boundary, especially in the PZ (Fig. 3 *B1*, *B3*, *C1*, and *C3* and *SI Appendix, Fig. S2*). The CG tumor barely deformed the posterior aspect of the PZ. During the shift in tumor morphology, the displacement fields redistributed and decreased globally between 23% and 31%, taking 0.3–0.6 y to recover during fingered growth. The urethra was displaced away from the tumor and also constricted (max. 0.44–0.75 mm), mainly in the segments circumvented by the fingered tumors.

The hydrostatic stress was compressive within the tumors, with lower values in the PZ (–0.75 kPa to –0.33 kPa; *SI Appendix, Fig. S2*). Around the tumors, the hydrostatic stress was compressive in the PZ, with a minimum of –0.08 kPa, and tensile in the CG, with a maximum of 0.10 kPa. Maximal tensile hydrostatic stress between 0.67 kPa and 0.88 kPa was obtained on the most constricted contours of the urethra and was accompanied by compressive values with minimum between –0.52 kPa and –0.38 kPa on the urethral contours in a perpendicular direction to the constriction. High tensile hydrostatic stress was also registered when tumors grew near the prostate external surface (up to 0.33 kPa in PZ and 0.48 kPa in CG). Far from the tumor, the hydrostatic stress was negligible.

The Von Mises stress ranged between 0.05 kPa and 0.25 kPa inside the tumor and it was higher around the convex side of the tumor borders, with values typically between 0.45 kPa and 0.60 kPa (*SI Appendix, Fig. S2*). Maximal values of the Von Mises stress were obtained in the healthy tissue between tumor branches (0.60–0.70 kPa), within intricate concave regions of fingered tumors (0.60–0.75 kPa), within and around the tumor near the border between the PZ and the CG (0.41–0.87 kPa), in healthy tissue compressed by the tumor against the prostate external surface (0.50–0.92 kPa), and on the contour of the most constricted segments of the urethra (0.75–1.27 kPa). The healthy tissue enclosed by tumor branches had Von Mises stress values within the intratumoral range. Massive tumors tended to build up greater values of the Von Mises stress around them. We observed that the Von Mises stress was slightly higher and the distortion rim was thicker in the CG.

**BPH Impedes PCa Growth.** After the deformational study of the prostate under the effect of BPH and PCa to calibrate our model, we analyzed how the patient’s history of BPH affected PCa growth. We ran two simulations: one without considering BPH and another including it. Fig. 4 depicts the growth of the tumor in both scenarios, while Fig. 5 shows the deformational states (*SI Appendix, Figs. S3* and *S4*). The geometry of the patient’s tumor was extracted from the same MR dataset used to obtain the prostate anatomy. We estimated the stress state



**Fig. 3.** Deformation of the prostate over 1 y produced by a tumor originated on basal PZ (A1–C1), apical PZ (A2–C2), and median CG (A3–C3). (A1–A3) Tumor growth over the original prostate geometry. (B1–B3) Length of the displacement field vector over original anatomy at  $t = 1$  y. The contour of the tumor is depicted with black curves. (C1–C3) Original and deformed geometries of the prostate at  $t = 1$  y.

of the prostate caused by years of developing BPH before PCa detection at the MR date,  $\sigma^0$ . This information was introduced as a prestress to compute the baseline hydrostatic and Von Mises stresses in the mechanotransductive coefficient adjusting tumor dynamics (*Materials and Methods*). Hence, for this purpose,  $\sigma = \sigma^0 + \sigma^1$ , where  $\sigma^1$  represents the additional stresses developed since the detection of PCa at MR date. All stress values quoted are for  $\sigma$ .

The simulation without BPH rendered similar results to the PZ tumor cases in the previous section. The patient's tumor was located in the left basal aspect of the PZ and had a volume of 0.51 cc. Initially, this tumor grew with massive morphology, early invading the CG and progressively developing two lobes of preferential growth in the anteroposterior direction. This geometry contributed to an early shift in morphology between  $t = 0.3$  y and  $t = 0.4$  y, during which tumor growth was minimally slowed down. Afterward, the tumor grew faster with

fingered morphology, invading all of the prostate side where it had originated from median to basal height in the craniocaudal direction.

The final volume of the tumor was 6.32 cc. The urethra was displaced anteriorly to the patient's right and the maximal constriction was 0.30 mm. The distribution and magnitude of the displacement, hydrostatic stress, and Von Mises stress fields were also analogous to their counterparts in the simulations for the PZ tumors in the previous section. The maximum total displacements were in the range 0.85–1.05 mm. The shape instability reduced displacements by 17.3% and it took until the end of the simulation for them to recover ( $t \approx 0.9$ ).

The inclusion of the patient's history of BPH produced major changes in tumor growth and the prostate deformational state. There was no shift in morphology in this simulation. Instead, the tumor grew as a continuous mass that progressively surrounded the CG in the anteroposterior direction at the basal level of the





36, 38). The mechanotransductive term to adjust tumor dynamics can be refined, for instance, by testing alternative formulations or differentiating how mechanical stress affects tumor mobility, proliferation, and apoptosis (25, 27, 33). To overcome the limitations of linear elasticity and obtain more accurate values of displacements and stresses future studies should explore a formulation accounting for geometric and material nonlinearities and investigate alternative constitutive equations for the prostate (28, 46, 54, 55). A poroelastic description of PCa growth would also help to study the effect of mechanical deformation on nutrient transport (26). This approach holds the potential to accurately investigate the heterogeneity of intratumoral metabolism or test patient-specific drug administration to the tumor in silico. To gain further knowledge on the mechanisms of PCa growth using our model, we believe it would be valuable to investigate the mathematical properties of the equations composing it, for example, analyzing solvability conditions, characterizing parameter spaces, and studying the well-posedness of the problem.

Finally, further validation using longitudinal series of clinical data is required to corroborate the BPH-induced mechanical inhibition of PCa. If validated, this mechanism may produce an important shift in the clinical management of BPH. Our results advise performing a comprehensive examination of the prostate in men without PCa before prescribing BPH treatments or drugs aimed at reducing the CG volume. Likewise, our results recommend a close follow-up of those patients with PCa receiving these sorts of BPH drugs. Otherwise, the prostate volumetric decrease could hazardously lower the mechanical stress fields created by BPH and hence reduce their mechanical restraint on PCa growth. Indeed, this could partially explain the higher proportion of more aggressive cancers in the finasteride arm of the Prostate Cancer Prevention Trial (56), arguably the most controversial result of this milestone study on chemoprevention of PCa (14, 18–20). Our research also calls for further courses of treatment in alleviating BPH symptoms while maintaining a significant stress level within the prostate for protection against PCa. In the future, we intend to address this issue by extending our model to incorporate the effect of drugs causing a prostate volume decrease, such as dutasteride and finasteride (1, 56). Additionally, we think that our model could be exploited to improve the current technologies for deformable registration of medical images (42) by providing a theory for the evolution of BPH and PCa. The resulting algorithms would be of much interest for several applications within the clinical management of PCa on a tissue-scale, patient-specific basis, such as the accurate follow-up of tumor growth during active surveillance or the precise planning of surgery and other treatments (39). Such technology could also help to detect PCa based on the deformation of the prostate boundaries and the internal borders between prostatic regions (44, 57). Additionally, it could guide biopsies to better stage and diagnose PCa. Comparing the predictions of the model with new images and clinical data could also help to discern whether the tumor is progressing toward a more malignant behavior and to make clinical decisions accordingly (39).

## Materials and Methods

**Patient Data.** Anonymized patient data were obtained from the public repository for PCa imaging data at the Initiative for Collaborative Computer Vision Benchmarking website ([i2cvb.github.io/](https://i2cvb.github.io/)) (58). Institutional review board approval and informed consent were not required for this study. We used the multiparametric MR image dataset obtained with a 3.0 Tesla Siemens Magnetom Trio TIM scanner. Patient cohort description and details on data acquisition have been previously described (59). For each biopsy-confirmed patient, this database includes T2-weighted MR images; dynamic contrast enhanced MR images; diffusion-weighted MR images; MR spectroscopic images; apparent diffusion coefficient maps; and the segmentations of the prostate, the PZ, the CG, and the tumor by an experienced radiolo-

gist. For this research, we selected a patient aged 54 y at MR date who had a large prostate of 52.81 cc harboring a localized tumor in the PZ.

**Mathematical Model.** Our modeling philosophy aims at developing tumor growth models based on key phenomena and featuring a limited number of representative parameters (30). This approach holds the potential for precise heterogeneous parameterization using available longitudinal clinical and imaging data from patients and has proved successful in reproducing and predicting various types of tumor growth (30, 32–34). As a preliminary step in this direction, here we developed a mechanically coupled model for patient-specific, organ-confined PCa growth and we leveraged it to perform a study on the mechanical interaction between PCa and BPH using literature data for parameter calibration.

Our model is described by the following equations:

$$\frac{\partial \phi}{\partial t} = M \left( D_\phi \Delta \phi - \frac{1}{\tau} \frac{dF(\phi)}{d\phi} + \chi s - A \phi \right) \quad [1]$$

$$\frac{\partial s}{\partial t} = D_s \Delta s + S - \delta \phi - \gamma_s s \quad [2]$$

$$\frac{\partial p}{\partial t} = D_p \Delta p + \alpha_h (1 - \phi) + \alpha_c \phi - \gamma_p p \quad [3]$$

$$\nabla \cdot \sigma = 0. \quad [4]$$

Eqs. 1–3 come from our previous model of organ-confined PCa growth (39, 40). Eq. 1 describes tumor dynamics using the phase-field method (41). The order parameter  $\phi$  takes values from 0 in healthy tissue to 1 within tumoral regions, showing a thin diffuse interface in between.  $F(\phi) = 16\phi^2(1-\phi)^2$  is a double-well potential, which enables the stable coexistence of healthy and cancerous tissue in our model. The last two terms in Eq. 1 describe nutrient-driven growth and apoptosis (i.e., programmed cell death), respectively. We consider that tumor growth is driven by a generic nutrient  $s$  that follows reaction–diffusion dynamics in Eq. 2. The reactive terms in Eq. 2 are the nutrient supply, the consumption of nutrient by the tumor, and a natural decay, respectively. Eq. 3 describes the reaction–diffusion dynamics of tissue PSA  $p$ , defined as the serum PSA concentration leaked to the bloodstream per unit volume of prostatic tissue (39). In Eq. 3, we consider that healthy tissue and cancerous tissue produce  $p$  at rates  $\alpha_h$  and  $\alpha_c$ , respectively, and the last term is a natural decay.

The mechanical stress fields generated by a growing tumor have been shown to slow down its dynamics (23–28). Previous clinically oriented, mechanically coupled models of tumor growth have incorporated this phenomenon by means of a coefficient that exponentially decreases cell mobility or net cell proliferation as mechanical stress fields intensify (30, 32–34). Following a similar strategy, in Eq. 1 we modeled the mechanically induced inhibition of tumor growth throughout coefficient  $M$ , which we define in Eq. 5. Mechanical stress can be decomposed as the sum of hydrostatic stress  $\sigma_h$  (defined in Eq. 6), which tends to change the volume of the stressed body, and deviatoric stress, which tends to distort it (46). The Von Mises stress  $\sigma_v$  (46) (defined in Eq. 7) is a good measure of the distortional strain energy around the tumor that has been widely adopted in mechanically coupled models (32–34). However,  $\sigma_v$  is insensitive to hydrostatic stress, which better describes the natural stress state within a growing region of tissue, such as a tumor or the CG developing BPH (23, 24, 26). Hydrostatic stress is also bound to increase globally during these growth processes because the prostate is a confined organ (1). Therefore, we propose to combine  $\sigma_v$  and the hydrostatic stress  $\sigma_h$  (46) in the definition of  $M$  following a similar approach to that of multiaxial stress-based failure criteria,

$$M = e^{-\beta_1(\sigma_v + \beta_2|\sigma_h|)}, \quad [5]$$

where

$$\sigma_h = \frac{1}{3} \sigma : \mathbf{I} = \frac{1}{3} (\sigma_{11} + \sigma_{22} + \sigma_{33}), \quad [6]$$

$$\sigma_v = \left( \sigma_{11}^2 + \sigma_{22}^2 + \sigma_{33}^2 - \sigma_{11}\sigma_{22} - \sigma_{22}\sigma_{33} - \sigma_{33}\sigma_{11} + 3 \left( \sigma_{12}^2 + \sigma_{23}^2 + \sigma_{13}^2 \right)^{1/2} \right)^{1/2}, \quad [7]$$

where  $\sigma_{ij}$  with  $i, j = 1, 2, 3$  are the components of the stress tensor  $\sigma$ ,  $\mathbf{I}$  is the second-order identity tensor, and  $\beta_1$  and  $\beta_2$  are constants that were adjusted to match the experimental and clinical results of previous studies on tumor growth (23–28, 32–34). We use the absolute value of the hydrostatic stress because both growth-induced compressive and tensile solid stresses have been shown to impede tumor dynamics at tissue scale in vivo (23, 24).

We assume that PCa and BPH show sufficiently slow rates (1) to neglect the inertial effects in the deformation of prostatic tissue. Hence, Eq. 4



describes mechanical equilibrium as a quasistatic process. Linear elasticity has been widely accepted to model mechanical equilibrium in living tissue subjected to slow processes over short time scales ( $t \sim 1$  y) (32–38, 46). Hence, this paradigm provides a simple mechanical framework to compute the stresses in Eq. 5 and perform our qualitative study on the mechanical influence of BPH on PCa. The prostate is an histologically heterogeneous organ: While the CG has a higher and denser stromal component, the PZ has more abundant glandular elements with sparsely interwoven smooth muscle (1, 5, 6). Benign enlargement of the prostate tends to make the CG denser and more compact (1, 7). Therefore, the CG is normally stiffer than the PZ in patients with PCa and/or BPH (42–45). The growing tumor induces an internal compressive hydrostatic stress and exerts outward forces acting on the tumor border (22, 24), so we modeled the tumor mass effect as

$$p_{\text{tumor}} = -\kappa\phi. \quad [8]$$

This assumes that  $\kappa$  is the magnitude of a constant compressive pressure, which is admissible over short simulation times (32–34, 36, 37). Volumetric growth due to BPH is usually modeled with an exponential function, but the slow growth rates justify a linear approximation over periods  $\Delta t \sim 1$ –10 y (51–53). Hence, we modeled BPH as a linearly growing pressure acting only on the CG,

$$p_{\text{BPH}} = -K\varrho \frac{g\Delta t}{V_{\text{MRI}}} \mathcal{H}_{\text{CG}}(\mathbf{x}), \quad [9]$$

where  $K$  is the bulk modulus,  $g$  is the estimated linear volumetric growth rate of the prostate due to BPH at MR date,  $V_{\text{MRI}}$  is the volume of the prostate as measured on T2-weighted MRI,  $\Delta t$  is the time passed since the date of MRI, and  $\mathcal{H}_{\text{CG}}(\mathbf{x})$  is a Heaviside function with value 1 in the CG and 0 elsewhere. The parameter  $\varrho$  adjusts the value of the pressure  $p_{\text{BPH}}$  to obtain the estimated CG growth rate. Taking all of the considerations above, we model prostatic tissue as a linear elastic, heterogeneous, isotropic material whose constitutive equation is given by

$$\sigma = \lambda (\nabla \cdot \mathbf{u}) \mathbf{I} + 2\mu \nabla^s \mathbf{u} - \kappa\phi \mathbf{I} - K\varrho \frac{g\Delta t}{V_{\text{MRI}}} \mathcal{H}_{\text{CG}}(\mathbf{x}) \mathbf{I}, \quad [10]$$

where  $\sigma$  is the stress tensor,  $\lambda$  and  $\mu$  are the Lamé constants,  $\mathbf{u}$  is the displacement vector, and  $\mathbf{x}$  is the position vector. The segmentation of CG and PZ was extracted from our patient's imaging data and mapped over the quadrature points to define heterogeneous material properties (Fig. 1).

We computed tumor volume  $V_\phi$  and serum PSA  $P$  as (39)

$$\begin{aligned} V_\phi &= \int_\Omega \phi d\Omega \\ P &= \int_\Omega p d\Omega, \end{aligned} \quad [11]$$

where  $\Omega$  is the prostate segmented on the patient's MR images.

Because we are focusing on localized PCa, we imposed zero-valued Dirichlet conditions for  $\phi$  all over the prostate boundary  $\partial\Omega$ . We set natural boundary conditions for  $s$  and  $p$ . The confinement of the prostate within the pelvic area (1) was modeled with Winkler-inspired boundary conditions on the external surface of the prostate, while free displacement was enabled along the urethra; i.e.,

$$\begin{aligned} \sigma \mathbf{n} &= -k_w \mathbf{u} && \text{in } \partial\Omega|_{\text{exterior}} \\ \sigma \mathbf{n} &= 0 && \text{in } \partial\Omega|_{\text{urethra}} \end{aligned} \quad [12]$$

where  $\mathbf{n}$  is the outer normal vector to  $\partial\Omega$  and  $k_w$  is constant.

The initial condition of the phase field,  $\phi_0$ , was an  $L^2$  projection of the tumor segmentation extracted from the patient's T2-weighted MR images and mapped over quadrature points or was artificially modeled with an  $L^2$ -projected hyperbolic tangent field. The initial conditions for  $s$  and  $p$  were approximated with linear functions based on  $\phi_0$  (40). Linear elasticity allows us to apply the principle of superposition and set zero-valued initial conditions for the displacements; i.e.,  $\mathbf{u}_0 = \mathbf{0}$ . This means that we are computing only the displacements produced since the MR date.

To study how BPH mechanically influences PCa growth, we estimated the stress state  $\sigma^0$  caused by the history of BPH before the detection of PCa at MR date and leveraged it as a prestress state to compute  $M$  in Eq. 1. Hence, for this purpose,  $\sigma = \sigma^0 + \sigma^1$ , where  $\sigma^1$  are the stresses developed since the detection of PCa at MR date. To estimate  $\sigma^0$ , we assumed that the volume of the patient's prostate was 20 cc at age 40 y (1, 3) and leveraged Eq. 4

with a negative value for  $g$  and  $\Delta t = 14$  y to approximate the undeformed, healthy state of our patient's prostate according to the standard anatomical features (1). Because we are using linear elasticity, it suffices to reverse the sign of the obtained displacements to yield  $\mathbf{u}^0$  and then approximate  $\sigma^0$  as

$$\sigma^0 = \lambda (\nabla \cdot \mathbf{u}^0) \mathbf{I} + 2\mu \nabla^s \mathbf{u}^0 - K\varrho \frac{g\Delta t^0}{V_{\text{MRI}}} \mathcal{H}_{\text{CG}}(\mathbf{x}), \quad [13]$$

where  $g$  is now positive and  $\Delta t^0 = 14$  y.

Parameter selection for Eqs. 1–3 has been previously discussed (39) and it is provided in Table 1. We set the values of the Young modulus of PZ and CG to  $E_{\text{PZ}} = 3$  kPa and  $E_{\text{CG}} = 6$  kPa, respectively (42–45). Because living soft tissues have a high content of water (34, 37, 42), we set the Poisson coefficients  $\nu_{\text{PZ}} = \nu_{\text{CG}} = 0.40$ . We empirically calibrated  $\beta_1 = 0.80$  1/kPa and  $\beta_2 = 1.50$  in agreement with the experimental and clinical observations reported in previous studies on tumor growth (23–28, 32–34). We selected  $\kappa = 2.50$  kPa to produce displacements in the order of 1 mm. The analysis of BPH and PCa deformation and the computation of the stress fields  $\sigma^0$  rendered  $g = 2.34$  cc/y,  $\varrho = 2.78$ , and  $k_w = 0.23$  kPa/mm. These values for  $g$  and  $\varrho$  reasonably agree with previous observations in the literature for our patient characteristics (1, 3, 50–53).

Finally, we acknowledge that linear elasticity is an acceptable simplification for studies featuring small strains and rotations and that the strain values in the simulations shown in Figs. 3–5 are somewhat outside the admissible range for linear elasticity in a few localized regions along the tumor interface. Additionally, the prestress  $\sigma^0$  is just a gross estimate of the stress state in the prostate due to the history of BPH previous to PCa detection at MR. However, we are using the stresses only to compute the mechanotransductive factor to adjust tumor dynamics. This study also features other major sources of uncertainty beyond linear elasticity, such as prostate and tumor segmentation, the PCa model itself, or the mechanical boundary conditions. Still, our predictive simulations still qualitatively reproduce the inhibiting effect on tumor growth caused by the tumor mass effect and a history of BPH before PCa detection. To verify the validity of the use of linear kinematic theory, we computed the symmetrical and skew-symmetrical components of the displacement gradient with respect to the initial configuration and determined that they were sufficiently small that their products were not substantial compared with the linear terms and, in particular, that the skew-symmetric components were negligible everywhere. Consequently, we feel confident that the computed results provide physically meaningful information.

**Numerical Methods.** Since tumor growth and BPH are modeled as quasistatic processes, we adopted a staggered approach to solve the equations in our mathematical model. We calculated tumor growth at every time step, but we updated the displacements only every two time steps. We performed spatial discretization by means of a standard isogeometric Bubnov–Galerkin approach using a 3D  $C^1$  quadratic nonuniform rational B-spline (NURBS) space (39, 40, 60–62). Temporal integration in the tumor growth problem was carried out with the generalized- $\alpha$  method (63, 64). This technique led to a nonlinear problem in each time step, which we linearized using the Newton–Raphson method. The resulting linear system was solved using the generalized minimal residual method (GMRES) algorithm (65) with a diagonal preconditioner. We also used the preconditioned GMRES algorithm to solve the quasistatic elastic problem. We chose a constant time step of

**Table 1. List of parameters in Eqs. 1–3**

Parameter	Notation	Value
Diffusivity of the phase field	$D_\phi$	200 mm <sup>2</sup> /y
Time scale for the phase field	$\tau$	0.01 y
Nutrient-induced tumor growth rate	$\chi$	600 L/(g · y)
Apoptosis rate	$A$	600 1/y
Nutrient diffusivity	$D_s$	5.47 · 10 <sup>3</sup> mm <sup>2</sup> /y
Nutrient supply	$S$	2.70 g/(L · d)
Nutrient consumption rate	$\delta$	2.75 g/(L · d)
Nutrient natural decay rate	$\gamma_s$	1,000 1/y
Tissue PSA diffusivity	$D_p$	200 mm <sup>2</sup> /y
Healthy tissue PSA production rate	$\alpha_h$	6.25 (ng/mL)/(cc · y)
Tumoral tissue PSA production rate	$\alpha_c$	$\alpha_c = 15\alpha_h$
Tissue PSA natural decay rate	$\gamma_p$	100 1/y

0.002 y and the parameters in the generalized- $\alpha$  method were set as in previous studies (40, 66).

**Visualization.** We used ParaView (67) to visualize and explore the results of our simulations. We used the isovolume  $\phi \geq 0.5$  to represent the tumor, which permitted us to study the evolution of PCa morphologies. From the  $C^1$ -continuous basis, we computed the stress fields pointwise. We identified regions of interest for the stress fields in the simulations and isolated them using appropriate geometric filters in ParaView. The stresses reported in the text correspond to the range of values that best describe the stress field in each region of interest.

**Construction of the Prostate Mesh.** Multiple methodologies permit the construction of solid anatomic NURBS models (68, 69). Because the geometries of the prostate and a solid torus are topologically equivalent, we leveraged a parametric mapping algorithm (70, 71) to deform a torus solid NURBS model to match with a patient-specific prostate surface model. We used 3DSlicer (72) to generate a triangular surface model of the prostate from the contours of the organ and the urethra drawn on the T2-weighted

MR images, using the provided prostate segmentation as guidance. The resulting surface was smoothed in MeshLab (73).

The original torus and prostate NURBS meshes were discretized with  $32 \times 32 \times 8$  elements along the toroidal direction, the cross-section circumferential direction, and the cross-section radial direction, respectively. We globally refined the prostate mesh to  $256 \times 256 \times 64$  elements, using standard knot insertion (61) to perform our simulations with a good level of accuracy.

**ACKNOWLEDGMENTS.** We thank Pablo Orosa-Iglesias (Universidad de Coruña, Spain) for his valuable help in the construction of the prostate mesh. We acknowledge the Centro de Supercomputación de Galicia (Santiago de Compostela, Spain) for providing high-performance computing resources that contributed to the results presented in this paper. G.L., P.D.-F., and H.G. were partially supported by the European Research Council (Contract 307201) and Xunta de Galicia (Consellería de Cultura, Educación e Ordenación Universitaria). G.L. and A.R. were partially supported by Fondazione Cariplo–Regione Lombardia through the project “Verso nuovi strumenti di simulazione super veloci ed accurati basati sull’analisi isogeometrica,” within the program RST–rafforzamento.

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